

A Depolymerizing Effect of the Luteinizing Hormone and Follicle Stimulating Hormone on the Neurotubules of the Hypothalamus-Hypophyseal Tract in the Rat

Cytoplasmic microtubules are made up of protein subunits which have the ability to change from isolated monomers to fibrous microtubules and vice versa. Much interest is being placed on the structural stability of microtubules. It is known that microtubules in neurons (neurotubules) are relatively more stable than those of other cell types. Disorganization of neurotubules have been reported after *in vitro* treatments with antimetabolic drugs¹, or elevated temperatures². Low temperatures, on the other hand, are effective to depolymerize microtubules in a variety of cells including amphibian neurons³.

In the present account we report a disorganization of neurotubules in nerve fibers of the hypothalamus-hypophyseal tract of the rat after *in vitro* treatments with follicle stimulating hormone (FSH), or luteinizing hormone (LH).

Male rats (120–130 g) were anesthetized with ether, decapitated and their hypothalami dissected out. Each hypothalamus was incubated in 1 ml of medium 199 (Difco). In the control group 0.1 ml of saline was added to the medium. The experimentals were added with 0.1 ml of LH (100 μ g/ml) or FSH (100 μ g/ml) in saline solution. The incubation was performed in a shaker in an atmosphere of 95% O₂, 5% CO₂ at 37°C for 6 h, according to WATANABE *et al.*⁴. The hypothalami were fixed in KARNOVSKY⁵, post fixed in OsO₄ (overnight), dehydrated in ethanol and embedded in Epon. Sections were stained with uranyl-lead.

The amount of microtubules per fibre was found to depend on the fibre diameter. On the basis of this observation, the cross sectioned axons of the hypothalamus-hypophyseal tract were classified into 3 groups: 1. 1–3 μ m; 2. 3–5 μ m; 3. 5–7 μ m. Counts of microtubules were performed in each group and the standard errors were small.

Both FSH and LH were found to produce a significant reduction of microtubules in all groups of fibres so far analyzed (Figure 1). FSH was more active (60% of reduction) than LH (30%). High power views of control fibres revealed that most of their microtubules kept the normal spatial pattern of subunits up to the 6th h of incubation (Figure 2). In the material incubated with LH (Figure 3) or FSH (Figure 4), most of the resistant microtubules were expressed by irregular aggregations of subunits (Figures 3 and 4). Images of '8' shaped microtubules were frequently seen in preparations treated with LH but not in the ones incubated with FSH (Figure 3).

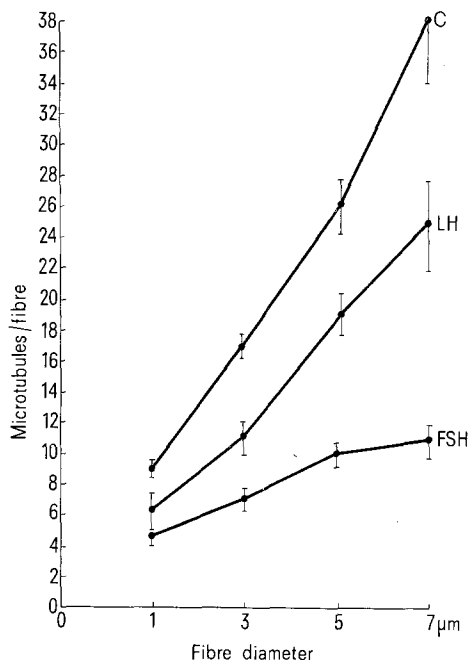


Fig. 1. Number of microtubules per cross sectioned fibres of the hypothalamus-hypophyseal tract (means \pm standard error of the means). Each value was obtained by counting the microtubules of over 1,300 fibres. Probability between Control (C) and LH treated fibres (LH) < 2%. Probability between C and FSH treated fibres (FSH) < 3%.

¹ H. WISNIEWSKI and R. D. TERRY, *Lab. Invest.* 17, 577 (1968).

² J. B. KIRKPATRICK, *J. Cell Biol.* 47, 384 (1970).

³ E. L. RODRIGUEZ-ECHANDIA, *J. Cell Biol.* 39, 491 (1968).

⁴ S. WATANABE, A. P. S. DHARIWAL and S. M. McCANN, *Endocrinology* 82, 674 (1968).

⁵ M. J. KARNOVSKY, *J. Cell Biol.* 27, 137 (1965).

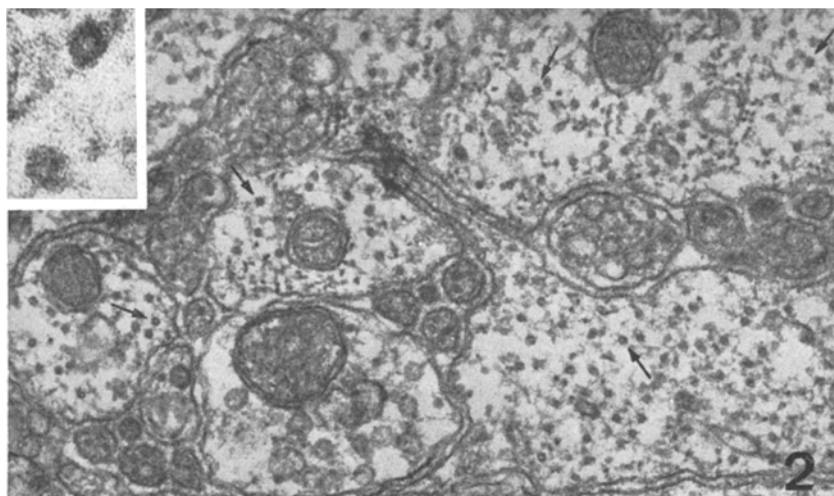


Fig. 2. Hypothalamus-hypophyseal tract incubated in medium 199 (control fibres). Microtubules (arrows) are abundant. The inset illustrates 2 microtubules at high magnification. $\times 40,000$. Inset: $\times 240,000$.

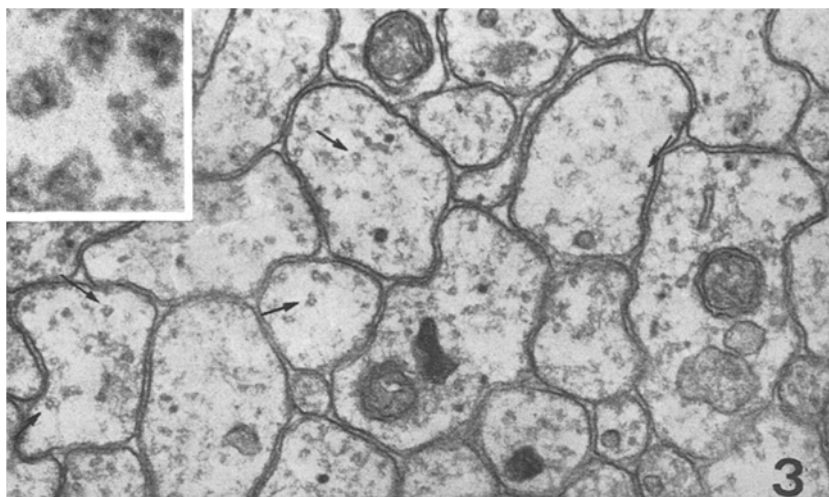


Fig. 3. LH treated hypothalamus. The fibres of hypothalamus-hypophyseal tract contain scanty microtubules (arrows). At high magnification disorganization of the microtubules subunits is apparent. $\times 40,000$. Inset: $\times 240,000$.

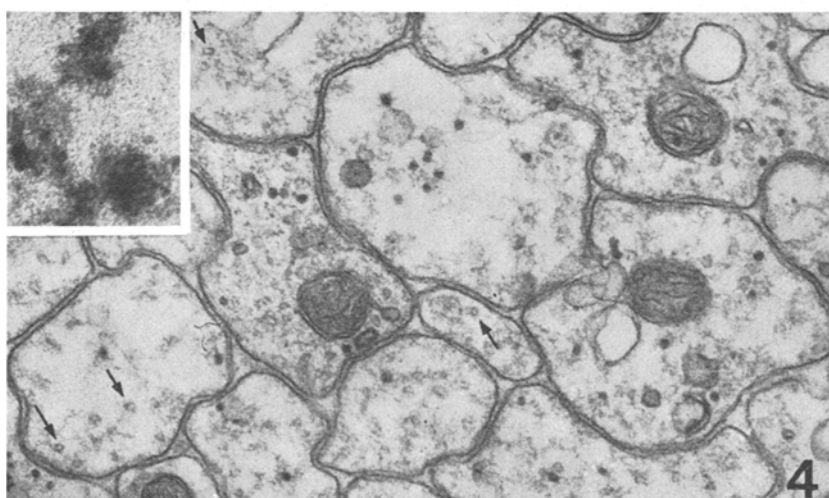


Fig. 4. FSH treated hypothalamus. Some FSH resistant microtubules are indicated by the arrows. The inset illustrates aggregation of microtubules subunits in zones presumably occupied by the pretreatment tubules. $\times 40,000$. Inset: $\times 240,000$.

It is known that tubulins (MW 120,000), the microtubule protein, polymerize into 12 or 13 subunits or protofibrils. Tubulins contain 2 mols of guanine nucleotides per 120,000 daltons⁶. The attractive interaction of the monomeric tubulins may be due to binding by hydrophobic residues⁷. Calcium ions are also essential for tubulin to polymerized into tubular structures⁸.

Several mechanisms have been proposed for microtubule depolymerization. Hydrostatic pressure and coldness would operate on the hydrophobic bounds joining the adjacent monomers⁷. Antimitotic drugs⁹, and divalent ions, e.g., nickel sulfate or copper sulfate, would replace a guanine nucleotide of tubulins dealing with a product which can be pelleted by ultracentrifugation¹⁰. Certainly, little can be said at present on the mechanism of microtubule disintegration by FSH or LH. However, gonadotrophins might act upon sulphhydryl groups of microtubules. The results are of interest, in view of the probable function of microtubules in translocating axoplasmic particulates such as neurosecretory granules¹¹ and synaptic vesicles¹².

In vivo experiments on various types of nerve fibres are to be done before conclusions can be reached on the significance of the reported lability of neuronal microtubules to gonadotrophins.

Résumé. Les neurotubules des fibres nerveuses du tract hypothalamo-hypophysaire du rat sont désorganisées après des traitements in vitro avec l'hormone lutéinisante et l'hormone folliculaire stimulante. Le mécanisme de dépolymérisation est discuté.

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⁶ M. L. SELANSKI, Proc. of the 24th American Ass. Neurochemistry Meeting 1971, p. 22.

⁷ M. L. SELANSKI and E. W. TAYLOR, J. Cell Biol. 34, 549 (1967).

⁸ L. E. ROTH and Y. SHIGENAKA, J. Ultrastruct. Res. 31, 356 (1969).

⁹ G. G. BORISY and E. W. TAYLOR, J. Cell Biol. 34, 525 (1967).

¹⁰ L. E. ROTH, J. Cell Biol. 34, 47 (1967).

¹¹ R. M. BERGLAND and R. M. TORACK, Expl. Cell Res. 54, 132 (1969).

¹² D. S. SMITH, U. JARLFORS and R. BERANEK, J. Cell Biol. 46, 199 (1970).